Vinylogous Hydroxamic Acids: 5-Lipoxygenase Inhibitors

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Abstract: Vinylogous hydroxamic acids were prepared as inhibitors of 5-lipoxygenase. The synthesis and preliminary SAR of these relatively unexplored compounds are described. These compounds are potent 5-lipoxygenase inhibitors which do not inhibit other enzymes of the arachidonic acid cascade.

Hydroxamic acids have recently received considerable attention as enzyme inhibitors, particularly of the enzyme 5-lipoxygenase, which is believed to play a key role in mediating certain inflammatory diseases. While hydroxamic acids (1) are well known and characterized compounds, the corresponding vinylogous hydroxamic acids, 3-(N-hydroxy-N-alkylamino)-2-propene-1-ones (VHAs, 2), represent an almost entirely unknown class of compounds. In analogy to vinylogous acids, VHAs would be expected to have chemical properties similar to those of hydroxamic acids. For example, VHAs could be able to form a bidentate chelate with transition metal ions, such as iron (III). The olefin in the VHAs allows additional substituents to be incorporated into the molecule. The greater diversity of structures than is possible with simple hydroxamic acids could confer additional interesting chemical and/or biological properties to the molecule.

$$R_1$$
 N OH R_1 R_2 R_3 R_4 R_2 R_3

To prepare these agents, we devised two synthetic routes (Methods A and B; Scheme 1). The synthesis of VHAs from appropriately substituted 1,3-dicarbonyl compounds (3)³ and hydroxylamines (R₄NHOH)⁴ was convenient (Method A) and provided a simple and inexpensive entry into this series.⁵ The reaction occurred

rapidly at 25° and proceeded best in protic solvents, particularly methanol, where the analytically pure VHAs usually crystallized directly from the reaction mixture. The reaction of 1,3-dicarbonyl compounds and hydroxylamines to give 2 also occurred in aprotic solvents such as chloroform, dichloromethane and THF.

Method B provided a convenient route to the VHAs by employing an exchange reaction of Me_2NH with hydroxylamines.⁶ The reaction of $E-3-(N_pN-dimethylamino)-2$ -propen-1-ones (4) with hydroxylamines to give 2 occurred rapidly in the presence of one equivalent of acid, usually p-toluenesulfonic or hydrochloric acid, in protic solvents. Methanol was the solvent of choice, as the VHAs generally precipitated directly from the reaction mixture in pure form. The reaction of 4 with hydroxylamines would not proceed to form 2 in the absence of acid.⁷

Scheme 1

$$R_1$$
 R_2
 R_3
 R_4
 R_3
 R_4
 R_3
 R_4
 R_5
 R_4
 R_5
 R_7
 R_8
 R_9
 R_9

(a) Method A: R₄NHOH, CH₃OH, 25°; (b) Method B: R₄NHOH, p-TsOH, CH₃OH, 25°.

Method A: $\frac{1-(4-\text{Fluorophenyl})-3-(N-\text{hydroxy-}N-\text{cyclohexyl})\text{amino-}2-\text{propen-}1-\text{one}}{2}$ (2b). A solution of 1-(4-fluorophenyl)-3-oxopropan-1-one (3, R₁ = 4-FC₆H₄; R₂ = R₃ = H)⁸ (3.32 g, 20 mmol) in 10 mL of methanol was heated to 40° and then treated at 40° with a solution of 2.30 g (20 mmol) of *N*-cyclohexylhydroxylamine in 20 mL of methanol. Crystals began to separate at once. The solution was allowed to cool to 25° during 1 hour. The product was filtered, washed with methanol and dried to afford 4.10 g (78%) of 2b as yellow crystals, mp 117° - 119°. ¹H NMR (CDCl₃): δ 7.79 (m, 2 H); 7.04 (t, 2 H); 7.01 (d, J = 6.4 Hz, 1 H); 5.34 (d, J = 6.4 Hz, 1 H); 3.66 (m, 1 H); 2.11 - 1.62 (m, 7 H); 1.41 - 1.18 (m, 3 H). The OH proton exchanged with D₂O. ¹³C NMR (C₆D₆): δ = 173.5, 165.9, 139.3, 129.6, 129.3, 115.0, 114.3, 69.3, 30.4, 26.5, 26.4. CIMS (NH₃): m/z = 264 (M + H⁺); 248 (M + H⁺ - O). IR (KBr pellet): 3500 cm⁻¹ (OH, weak); 1620 cm⁻¹ (C=O, strong); 1590 cm⁻¹ (C=C, strong). UV (EtOH): $\lambda_{\text{max}} = 362$ nm (log e = 4.29). Analysis: Calc'd for C₁₅H₁₈FNO₂: C 68.42%; H 6.89%; N 5.32%; Found C 68.59%; H 6.80%; N 5.13%.

Method B: 1-(4-Fluorophenyl)-3-(N-hydroxy-N-benzyl)amino-2-propen-1-one (2d). N-Benzyl-hydroxylamine (0.616 g, 5.0 mmol) and p-TsOH • H₂O (0.856 g, 4.5 mmol) were dissolved in 4 mL of 9:1 methanol:water and 0.966 g (5.0 mmol) of E-1-(4-fluorophenyl)-3-(N,N-dimethylamino)-2-propen-1-one (4, R₁ = 4-FC₆H₄; R₂ = R₃ = H)⁹ was added at 25°. The mixture was kept at 25° for 1 h, then cooled to -20°. Scratching induced crystal growth which was allowed to proceed for 20 min. The crystals were then filtered, washed with cold 9:1 methanol:water and dried to provide 0.433 g (34%) of 2d as yellow crystals, mp 98° - 100°. ¹H NMR (CDCl₃): δ 8.04 (d, J = 6.1 Hz, 1 H); 7.82 (m, 2 H); 7.44 - 7.23 (m, 5 H); 7.03 (t, 2 H); 4.99 (s, 2 H); 4.82 (d, J = 6.1 Hz, 1 H). The OH proton exchanged with D₂O. ¹³C NMR (CDCl₃): δ = 174.9, 165.7, 153.7, 133.8, 131.2, 129.3,

128.3, 126.4, 125.9, 114.9, 92.2, 63.9. CIMS (CH₄): m/z = 272 (M + H⁺); 256 (M + H⁺ - O). IR (KBr pellet): 3500 cm⁻¹ (OH, weak); 1620 cm⁻¹ (C=O, strong); 1590 cm⁻¹ (C=C, strong). UV (EtOH): $\lambda_{max} = 365$ nm (log e = 4.27). Analysis: Calc'd for C₁₆H₁₄FNO₂: C 70.84%; H 5.20%; N 5.16%; Found: C 70.58%; H 5.04%; N 5.03%.

The VHAs were examined for their ability to inhibit various enzymes in the arachidonic acid cascade thought to play a role in inflammatory diseases. They were found as a class to be inactive as inhibitors of cyclooxgenase (bovine seminal vesicles, IC_{50} generally > 750 μ M)¹⁰ and PLA₂ (Croatalus adamanteus, IC_{50} generally > 1mM).¹¹ As anticipated (Table 1), they were active as inhibitors of 5-lipoxygenase (rat basophilic leukemia cells).¹² A comparison of the *in vitro* activities of selected VHAs and some standard drugs is given in Table 2. The hydroxamic acid A64077 (Zileuton, 5)¹³ was chosen as a selective 5-lipoxygenase inhibitor, while phenidone (6) was selected as a dual 5-LO/CO inhibitor.

Table 1: Physical Data and 5-Lipoxygenase Inhibitory Activity of VHAs (2)

Entry	<u>R₁</u>	R ₂	<u>R3</u>	R ₄	R ₅	5-LO, IC ₅₀	Yield ^a	Method	mp, °Cb
2a	$4-FC_6H_4$	H	H	CH ₃	H	5.0 μΜ	20%	В	81 - 83
2b	4-FC ₆ H ₄	H	Н	c-C ₆ H ₁₁	Н	$0.27~\mu M$	78%	Α	117 - 119
2c	4-FC ₆ H ₄	H	H	C ₆ H ₅	Н	$0.20~\mu M$	50%	Α	157 - 159
2d	4-FC ₆ H ₄	H	H	CH ₂ C ₆ H ₅	Н	$0.15\mu M$	34%	В	98 - 100
2e	4-FC ₆ H ₄	H	Н	CH ₂ (4-Ph)C ₆ H ₄	Н	$3.0\mu M$	72%	Α	159 - 161
2f	4-FC ₆ H ₄	CH ₃	Н	CH ₂ C ₆ H ₅	Н	$0.30~\mu M$	30%	Α	198 - 200
2g	4-FC ₆ H ₄	H	CH ₃	CH ₂ C ₆ H ₅	Н	$0.25~\mu M$	14%	Α	95 - 97
2h	4-FC ₆ H ₄	H	Н	CH ₂ C ₆ H ₅	COCH ₃	$> 25 \mu M$	44%	C	115 - 117
2i	4-FC ₆ H ₄	H	H	CH ₂ C ₆ H ₅	CH ₃	$> 25 \mu M$	45%	Ad	76 – 78
2j	4-PhC ₆ H ₄	H	H	CH ₃	H	0.06 µM	65%	В	143 - 145

^a All yields reported are for purified products. ^b All compounds gave satisfactory ¹H NMR, CIMS, and elemental analyses. ^c Prepared by the acetylation of **2d** with Ac_2O (1.1 eq) and Et_3N (1.3 eq) in CH_2Cl_2 at 20 °C for 2 h.14 d Prepared from N-methoxybenzylamine.15,16

From the results in Table 1, it may be noted that the NOH moiety is critical for lipoxygenase inhibition. Acylation (2h, $R_5 = COCH_3$) or alkylation (2i, $R_5 = CH_3$) of the hydroxylamine oxygen results in the complete loss of inhibitory activity. This may be due to the reversal of the olefin geometry from Z (2a - d, $J \approx 6$ Hz) to E (2h and 2i, $J \approx 12$ to 18 Hz). The substitution of various groups on the hydroxylamine nitrogen (R_4) appears to

be well tolerated, with the *in vitro* potency increasing as R₄ becomes more lipophilic. Likewise, increasing the lipophilicity of R₁ also affords increased *in vitro* potency (2j vs. 2a). An extreme increase in lipophilicity, however, is detrimental and enzyme inhibition is decreased (2e).¹⁷ The loss of potency observed with 2e may also be attributed, wholly or in part, to the increased steric demands of the substituent R₄ in 2e. Placing substituents on the olefin (2f and 2g) results in a slight loss of potency *in vitro*. Studies are currently in progress to examine the biological properties of the VHAs *in vivo* and to determine the mechanism by which they inhibit lipoxygenase.

Table 2: In Vitro Profiles of Standard Drugs and Selected VHAs (2)

Compound	PLA_2^a	CO_p	5-LO ^c
	IC_{50}^d	IC_{50}^{d}	IC ₅₀ d
A64077	> 1000	> 750	0.14
Phenidone	> 1000	6.9	0.48
2 b	> 1000	> 750	0.27
2 d	> 1000	> 750	0.15
2 h	> 1000	> 750	> 25
2i	> 1000	> 750	> 25

^a PLA₂ inhibition. ^b Cyclooxygenase inhibition. ^c 5-Lipoxygenase inhibition. ^d Values listed in μM concentration; averages of two or more determinations.

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